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Cytotoxicity of curcumin-loaded nanoparticles based on amphiphilic poly-N-vinylpyrrolidone derivatives in 2D and 3D in vitro models of human ovarian adenocarcinoma

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ABSTRACT

BACKGROUND: Nanocarriers based on biocompatible polymers are a promising delivery tool for biologically active substances and drugs, in particular antitumor agents. Curcumin, a polyphenol, is known to possess pleiotropic therapeutic effects, including antitumor activity. The antitumor potential of curcumin has been shown in various tumor types, including ovarian adenocarcinoma. However, its lipophilic properties and very low bioavailability limits its use. Incorporating curcumin into nanocarriers enhances its delivery options and expands its potential as an antitumor agent.

AIM: To produce curcumin-loaded polymeric nanoparticles based on amphiphilic poly-N-vinylpyrrolidone derivatives and its copolymers with acrylic acid, explore their accumulation in the tumor cells; evaluate *in vitro* cytotoxicity in 2D (monolayer cell culture) and 3D (tumor spheroids) models of human ovarian adenocarcinoma.

MATERIALS AND METHODS: The polymers of the amphiphilic poly-N-vinylpyrrolidone derivatives and its copolymers with acrylic acid were obtained using radical polymerization. Emulsion method was used to obtain polymeric nanoparticles. Accumulation of nanoparticles in tumor cells was assessed using flow cytometry (for monolayer culture) or fluorimetric analysis (for spheroids). Cytotoxicity was studied in 2D and 3D models obtained of the human ovarian adenocarcinoma cell line OVCAR-3 using 3-4,5-dimethylthiazol-2-yl-2,5-diphenyltetrazolium bromide (MTT assay).

RESULTS: The effective accumulation of curcumin-loaded polymeric nanoparticles in both monolayer culture cells and tumor spheroids was demonstrated. Curcumin-loaded nanoparticles exhibited high-level cytotoxicity in the 2D model of human ovarian adenocarcinoma cells OVCAR-3 (IC50 up to 137±9 µg/mL) and a moderate, although significant cytotoxic effect in a 3D *in vitro* model. Meanwhile, nanoparticles not loaded with curcumin did not show any cytotoxic activity regardless of their composition or of the additional modification, i.e. with the use of maleimide functional groups.

CONCLUSION: These data can provide a foundation for further studies to assess the safety and *in vivo* antitumor activity of curcumin-loaded nanoparticles based on amphiphilic poly-N-vinylpyrrolidone derivatives.

Keywords: curcumin; amphiphilic poly-N-vinylpyrrolidone derivatives; nanoparticles; human ovarian adenocarcinoma; OVCAR-3 cell line.

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Исследование цитотоксичности наночастиц на основе амфифильных производных поли-N-винилпирролидона, загруженных куркумином, на 2D- и 3D-моделях *in vitro* аденокарциномы яичника человека

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АННОТАЦИЯ

Обоснование. Наноносители на основе биосовместимых полимеров перспективны для доставки биологически активных веществ и лекарств, в частности противоопухолевых агентов. Известно, что полифенол куркумин обладает плейотропными терапевтическими эффектами, включая противоопухолевую активность. Противоопухолевый потенциал куркумина показан на различных типах опухолей, в том числе на аденокарциноме яичника. Однако его применение ограничено его липофильной природой и очень низкой биодоступностью. Инкапсулирование куркумина в наноносители позволяет расширить спектр его применения, а также изучить возможность использования в качестве противоопухолевого препарата.

Цель. Получение полимерных наночастиц на основе амфифильных производных поли-N-винилпирролидона и его сополимеров с акриловой кислотой, загруженных куркумином; изучение их накопления в опухолевых клетках; оценка цитотоксичности *in vitro* в 2D- (монослойная культура клеток) и 3D-моделях (опухолевые сфероиды) на основе клеточной линии аденокарциномы яичника человека.

Материалы и методы. Полимеры на основе амфифильных производных поли-N-винилпирролидона и его сополимеров с акриловой кислотой получены радикальной полимеризацией. Полимерные наночастицы получали эмульсионным методом. Накопление наночастиц в опухолевых клетках изучали с помощью проточной цитометрии (монослойная культура) или флуориметрии (сфероиды). Цитотоксичность исследовали с помощью МТТ-теста на 2D- и 3D-моделях на основе клеточной линии аденокарциномы яичника человека OVCAR-3.

Результаты. Показано эффективное накопление полимерных наночастиц, загруженных куркумином, как в клетках монослойной культуры, так и в опухолевых сфероидах. Наночастицы, загруженные куркумином, показали высокий уровень цитотоксичности для клеток аденокарциномы яичника человека OVCAR-3 в 2D-модели (IC50 до 137±9 мкг/мл) и умеренный, но достаточно очевидный цитотоксический эффект на 3D-модели *in vitro*. При этом у всех образцов наночастиц, не загруженных куркумином, цитотоксическая активность отсутствовала независимо от их состава или наличия дополнительной модификации функциональными малеимидными группами.

Заключение. Полученные данные могут лечь в основу дальнейших исследований по безопасности и противоопухолевой активности *in vivo* наночастиц на основе амфифильных производных поли-N-винилпирролидона, загруженных куркумином.

Ключевые слова: куркумин; амфифильные производные поли-N-винилпирролидона; наночастицы; аденокарцинома яичника человека; клеточная линия OVCAR-3.

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BACKGROUND

In contemporary medicine, nanocarriers play an important role, providing opportunities for targeted delivery of anticancer agents directly to tumor cells, thus significantly reducing side effects and increasing their therapeutic effect. In particular, the incorporation of lipophilic anticancer agents in nanocarriers enhances their efficacy due to improved solubility, selectivity, and prolonged release [1].

Currently. polymeric nanocarriers, especially nanoparticles, are a promising tool, demonstrating high stability, low toxicity, and ability to efficiently incorporate lipophilic drugs [2]. Some micellar nanocarriers loaded with anticancer agents have already been approved for marketing or are currently being studied in clinical trials [3]. Polymeric nanoparticles based on amphiphilic block copolymers, enabling various structural and functional modifications of the formed delivery system, are currently under active research [4, 5]. Promising polymers include biocompatible and biodegradable poly-N-vinylpyrrolidone (PVP), which may be used to produce highly stable polymeric nanoscale particles [6]. For example, nanoparticles produced from amphiphilic PVP derivatives are stable in the presence of blood serum; they do not significantly affect blood cell function and complement activation, and have no hemolytic or inflammatory effects [7]. The previous studies demonstrated that such nanoparticles based on amphiphilic PVP derivatives may be loaded with Indometacin, the antiinflammatory drug [8], and bortezomib, the anticancer drug [9].

Curcumin, a polyphenolic pigment derived from the Curcuma longa rhizome, was shown to have a significant impact on various intracellular signaling pathways. In addition to its antitumor properties, curcumin is known possess anti-inflammatory, immunomodulatory, to neuroprotective, hepatoprotective, and antiviral effects [10]. Due to its lipophilic properties, curcumin is frequently used as a model drug in the development of novel delivery systems. Like most antitumor agents used in clinical practice, curcumin possesses antitumor activity and is easily detected by fluorescence. The antitumor potential of curcumin has been shown in various tumor types, particularly human ovarian adenocarcinoma [11, 12]. However, its low chemical stability and bioavailability of curcumin due to its lipophilic properties limit its therapeutic use [13]. Incorporating curcumin into nanocarriers may result in the production of stable and effective dosage forms of curcumin, thereby increasing its efficacy and expanding its application range, including its use as an additional antitumor therapy.

The aim of study was to produce curcumin-loaded polymeric nanoparticles based on amphiphilic PVP derivatives and its copolymers with acrylic acid, as well as to explore their accumulation in tumor cells and evaluate *in vitro* cytotoxicity in 2D (monolayer cell culture) and 3D (multicellular spheroids) models based on the OVCAR-3 human ovarian adenocarcinoma cell line.

The study demonstrated the cytotoxic activity of curcumin-loaded polymeric nanoparticles based on amphiphilic poly-N-vinylpyrrolidone derivatives and its copolymers with acrylic acid against human ovarian adenocarcinoma cells in two *in vitro* models. In addition, it was shown that nanoparticle surface modification with maleimide functional groups did not result in cytotoxicity. Such modified nanoparticles may be used in the future for covalent crosslinking with protein molecules (ligands) for targeted delivery to cancer cells. The obtained data may provide a basis for further studies to assess the safety and *in vivo* antitumor activity of curcumin-loaded nanoparticles based on amphiphilic PVP derivatives or other lipophilic drugs with antitumor activity.

MATERIALS AND METHODS

Study design

The study design involves three stages.

Stage 1 was to synthetize polymers based on amphiphilic PVP derivatives to produce polymeric nanoparticles with incorporated curcumin.

Stage 2 was to evaluate the accumulation efficiency of the nanoparticles by OVCAR-3 tumor cells (*in vitro* 2D and 3D models).

Stage 3 was to evaluate the cytotoxic activity of curcumin-loaded nanoparticles in 2D and 3D models based on *in vitro* OVCAR-3 tumor cells.

Study setting and duration

The study was conducted at the Shemyakin and Ovchinnikov Institute of Bioorganic Chemistry of the Russian Academy of Sciences, Mendeleev Russian University of Chemical Technology, and Lomonosov Moscow State University. The study lasted from March 2024 to September 2024.

Description of research methodology

Synthesis of polymers based on amphiphilic PVP derivatives

Two types of polymers representing amphiphilic PVP derivatives with a molecular weight of 3 kDa were synthesized: Amph-PVP (PVP with one terminal n-octadecyl fragment) and Amph-PVP-AA (copolymer of N-vinylpyrrolidone and acrylic acid with one terminal n-octadecyl fragment). The synthesis of Amph-PVP and Amph-PVP-AA proceeded as follows: 0.115 g (accurately weighed) of 2,2'-azobisisobutyronitrile (AIBN) and 0.429 g (accurately weighed) of octadecyl mercaptan were added to a 250 mL round bottom flask. This was followed by the addition of a solution of 10.7 mL of N-vinylpyrrolidone in 40 mL of 1,4-dioxane. For Amph-PVP-AA polymer, 350 µL of acrylic acid was also added to the system. The reaction mixture was mixed for 5 min until all components were completely dissolved. The resulting solution was allowed to stand for 1 h at room temperature. Subsequently, 100 mL of distilled water was added. Then, the resulting polymer mixture was distilled using a rotary evaporator (Hei-Vap Value Digital, Heidolph Instruments, Germany) to remove 1,4-dioxane. The prepared mixture was dialyzed against water (Slide-A-Lyzer™ Dialysis Flask, 1K MWCO, Thermo Scientific, USA) for four days and then lyophilized (Alpha 1-4 LD plus, Martin Christ, Germany). Functional analysis (potentiometric titration) was used to determine the molecular weight of the polymers.

Modification of Amph-PVP-AA polymers with maleimide groups

A 0.5 g suspension of Amph-PVP-AA was dispersed in 25 mL of water. The modification of Amph-PVP-AA involved preparation of two solutions: a 1 mg solution of 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide in 2 mL of water and a 16 mg solution of N-hydroxysuccinimide in 3 mL of water. These solutions were left to stir for 15 min before being added to the nanoparticle solution. A 4 mg (accurately weighed) of 1-(2-aminoethyl)maleimide was dissolved in 5 mL of water and subsequently added to the nanoparticle solution. Then, the reaction mixture was mixed for 24 h. The modified nanoparticles were then dialyzed against water, frozen, and lyophilized. The finished powder consisted of nanoparticles that could be dispersed in water or phosphate-salt buffer (pH=7.4) to produce a stable nanoparticle suspension.

Preparation of nanoparticles from curcumin-loaded amphiphilic PVP derivatives (Amph-PVP-Cur)

The Amph-PVP-Cur nanoparticles were produced by emulsion method, whereby 0.1 g of polymer was dispersed in 20 mL of water, and 0.0014 g of curcumin was dissolved in 5 mL of acetone. Then, the polymer solution was ultrasonicated for 5 min while cooling. Following the homogenization process, the curcumin solution was introduced to the polymer solution and subjected to additional ultrasonic treatment for another 5 min. The acetone was then extracted using a rotary evaporator (Hei-Vap Value Digital, Heidolph Instruments, Germany). The suspension was centrifuged (Sigma 4-5 L, Germany) to separate unincluded curcumin. The supernatant was then lyophilized. The finished powder consisted of nanoparticles, which could be dispersed in water or phosphate-salt buffer to produce a stable particle suspension.

Preparation of nanoparticles from curcumin-loaded amphiphilic PVP derivatives modified with acrylic acid and maleimide groups (Amph-PVP-AA-Mal-Cur)

The Amph-PVP-AA-Mal-Cur nanoparticles were produced by emulsion method, whereby 0.5 g of polymer

was dispersed in 25 mL of water, and 0.002 g of curcumin was dissolved in 5 mL of acetone. Then, the polymer solution was ultrasonicated for 5 min while cooling. Following the homogenization process, the curcumin solution was introduced to the polymer solution and subjected to additional ultrasonic treatment for another 5 min. The acetone was then extracted using a rotary evaporator (Hei-Vap Value Digital, Heidolph Instruments, Germany). The suspension was centrifuged (Sigma 4-5 L, Germany) to separate unincluded curcumin. The supernatant was then collected. The modification of nanoparticles involved preparation of two solutions: a 1 mg solution of 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide in 2 mL of water and a 16 mg solution of N-hydroxysuccinimide in 3 mL of water. These solutions were left to stir for 15 min before being added to the nanoparticle solution. A 4 mg (accurately weighed) of 1-(2-aminoethyl)maleimide was dissolved in 5 mL of water and subsequently added to the nanoparticle solution. Then, the reaction mixture was stirred for 24 h. The modified nanoparticles were then dialyzed against water, frozen, and lyophilized. The finished powder consisted of nanoparticles that could be dispersed in water or phosphate-salt buffer to produce a stable nanoparticle suspension. Particle sizes were determined by dynamic light scattering (Zetasizer Nano ZS, Marvern). The average size of the nanoparticles was in the range up to 350 nm.

The incorporation efficiency of curcumin into nanoparticles was calculated by the ratio of total curcumin content in the particles to the total amount of loaded curcumin. The release profile of curcumin from the nanoparticles was investigated over 24 h as previously described [14]. Briefly, the nanoparticles were suspended in phosphate-salt buffer (pH 7.4), after which the suspension was centrifuged (9000 g, 15 min), and the amount of released curcumin in the buffer was determined spectrophotometrically at 425 nm. Aliquots of samples were collected at 0.5, 1, 2, 6, 12, 16, and 24 h.

Cell cultivation

The OVCAR-3 human ovarian adenocarcinoma cell line (American Type Culture Collection, ATCC, USA, Cat. No. NTV-161) was provided by Viktor. V. Tatarsky, Cand. Sci. (Biology) (Institute of Gene Biology, Russian Academy of Sciences). The cells were cultivated in RPMI-1640 medium, which was enriched with 10% fetal bovine serum (FBS). The cells were cultured in a CO_2 incubator (N-BIOTEK *NB-203*, South Korea) in the gas-air medium containing 5% CO_2 at 37°C. The cell growth was monitored daily, using a light inverted microscope (Reichert Microstar 1820E, Germany).

Preparation of multicellular spheroids

Multicellular spheroids were derived directly from monolayer OVCAR-3 cell culture using the method that was developed by the authors earlier [14]. The cell suspension was added to a 96-well plate (SPL Lifesciences, Korea) at the rate of 7500 cells per well (in 100 μ L of RPMI-1640 nutrient medium, 10% FBS). Then, the plate was placed in a CO₂ incubator for 24 h. Subsequently, the medium was replaced with 100 μ L of fresh medium (with 10% FBS) containing 40 μ M of synthetic cyclic RGD peptide (cyclo-RGDfK(TPP). The plate was transferred to a CO₂ incubator where spontaneous aggregation of cells occurred within 72 h, forming multicellular spheroids. The size of the spheroids was determined by light microscopy. The average spheroid size was 150 nm.

In vitro study of polymeric nanoparticle accumulation efficiency

Flow cytometry

A flow cytometry method using a fluorescenceactivated flow cytometer (BD FACSCalibur, USA) with BD CellQuest software was used to quantitatively assess the effectiveness of polymeric nanoparticle accumulation in OVCAR-3 monolayer culture cells. Cells were seeded in a 24-well plate (50,000 cells per well) and incubated for 24 h (37°C, 5% CO₂). Then, the old culture medium was removed and fresh RPMI-1640 medium containing the suspension of curcumin-loaded polymeric nanoparticles (0.5 mg/mL, Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur samples, respectively) was added. The cells were then incubated with the samples for 5, 30, and 60 min, after which the unbound nanoparticles were removed by washing three times with phosphate-buffered saline (pH 7.4). The samples were analyzed at a wavelength of 488 nm. Flow cytometry data were expressed as the average fluorescence intensity divided by the background intensity of the control group (untreated cells).

Fluorimetry

The accumulation of polymeric nanoparticles in spheroid cells was quantitatively assessed by fluorimetric analysis. The culture medium was replaced with a fresh medium containing a 0.5 mg/mL suspension of the polymeric nanoparticles Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur and incubated (37 °C, 5% CO₂) with spheroids for 1, 3, and 7 h, respectively. Then, the spheroids were washed three times with PBS (pH 7.4) to remove unbound nanoparticles, and the average fluorescence level was measured (Promega GloMax-Multi detection system, USA) at a wavelength of 425 nm (Cur λ ex=571 nm, λ em=467 nm). Absorbance data were expressed as the percentage of nanoparticle fluorescence of the baseline solution.

In vitro study of the cytotoxicity of polymeric nanoparticles

The cytotoxicity of the nanoparticles was evaluated using 3-4,5-dimethylthiazol-2-yl-2,5-diphenyltetrazolium bromide (MTT assay). For monolayer culture (2D *in vitro* model), cells were dispersed in a 96-well culture plate

(7500 cells per well) and incubated in an incubator (5% CO2, 37 °C) for 24 h. For spheroid culture (3D in vitro model), a plate with pre-prepared spheroids was used. Then, the medium was removed and a fresh RPMI-1640 medium (10% FBS) containing nanoparticle samples at different concentrations (1, 10, 50, 100 and 500 µg/mL) was added to the cells or spheroids and incubated for 24 and 48 h. After incubation, cells or spheroids were treated with 0.05% MTT reagent solution in RPMI-1640 (without FBS) and left for 3 h. Then, the medium was removed and replaced with dimethyl sulfoxide (100 µL per well) and adsorption was measured using a Multiskan FC reader (Thermo Scientific, USA) at a wavelength of 540 nm. The semi-inhibitory concentration (IC50) was defined as the sample concentration that resulted in 50% growth inhibition of the cells. Monolayer cell culture and spheroids without nanoparticles were used as control (100% viable cells). The results of the MTT assay were processed using the GraphPad Prism software (USA).

Statistical analysis

All data were normally distributed and expressed as mean or mean±standard deviation. Two-way analysis of variance (ANOVA) followed by Sidak's multiple comparisons test was used to statistically analyze the flow cytofluorimetry and fluorimetry results. For the MTT assay results, ANOVA followed by Tukey's multiple comparison test was used for analysis. All experiments were performed in triplicate. The collected data were processed using GraphPad Prism software (GraphPad Software Inc., USA) and were found to be significantly different at p < 0.05.

RESULTS

Preparation of amphiphilic polymers and nanoparticles on their basis

A series of variants of amphiphilic N-vinylpyrrolidone polymers with varying hydrophilic composition were derived. In the first variant (Amph-PVP), the watersoluble block was represented by a N-vinylpyrrolidone polymer, and in the second variant (Amph-PVP-AA), it was represented by a copolymer of N-vinylpyrrolidone and acrylic acid. The incorporation of acrylic acid enables the introduction of 1% to 5% functional carboxyl groups, which may be used for subsequent surface modification of nanocarriers derived from these polymers [15]. In this study, the carboxyl group of acrylic acid was used to introduce maleimide functional groups into the polymer structure, thereby enabling the subsequent conjugation of a targeted anticancer agent under mild conditions. The hydrophilic segment, comprising copolymers of N-vinylpyrrolidone and acrylic acid, underwent modification with 1-(2-aminoethyl)maleimide in the presence of EDS/NHS as an intermediate. Subsequent to

this modification, amidation was initiated, resulting in the formation of the Amph-PVP-AA-Mal polymer (Fig. 1).

The hydrophobic site in both polymer variants was represented by an n-octadecyl moiety. The Amph-PVP, Amph-PVP-AA, and Amph-PVP-AA-Mal polymers were prepared by radical polymerization in the presence of an initiator and a chain growth regulator. This method allowed for the control of the molecular weight of the resulting polymers, which was 3 kDA for both polymer variants in this study. The structural and compositional characteristics of the polymers were determined and confirmed by several physicochemical analysis methods, including infrared spectroscopy, nuclear magnetic resonance spectroscopy, functional analysis, elemental analysis, and vapor pressure osmometry, as previously reported in our studies [15, 16].

The presence of both hydrophilic and hydrophobic fragments in the structure of the synthesized polymers promotes their self-assembly in aqueous media at

concentrations above a certain critical aggregation concentration with the formation of nanoscale associates of the "hydrophobic core-hydrophilic shell" type. Curcumin, as a poorly water-soluble substance, was loaded into the core of such particles through hydrophobic interactions with n-alkyl fragments of polymers by emulsion formation followed by solvent distillation. As a result, two types of nanoscale aggregates (Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur) were produced. The characteristics of the resulting particles are shown in Table 1.

The efficiency of curcumin incorporation into the samples ranged from 93% to 95%. The profile of curcumin release from nanoparticles was investigated over a 24-h period with time intervals of 0.5, 1, 2, 6, 12, 16, and 24 h (see Fig. 2).

A biphasic curcumin release profile was observed for Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur nanoparticles. During the first 30 min, 11% and 14% of the curcumin was released from the Amph-PVP-Cur



Scheme of the radical polymerization of N-vinylpyrrolidone in the presence of N-octadecyl mercaptan as a chain transfer agent



Scheme of the radical polymerization of N-vinylpyrrolidone and acrylic acid in the presence of N-octadecyl mercaptan



Fig. 1. Synthesis of amphiphilic derivatives of poly-N-vinylpyrrolidone for subsequent production of modified polymeric nanoparticles with curcumin.

In vitro study of polymeric nanoparticle accumulation in tumor cells

The accumulation efficiency of curcumin-loaded polymeric nanoparticles, in particular, Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur samples, was investigated by flow cytometry and fluorimetry (Fig. 3).

For monolayer OVCAR-3 cell culture (2D *in vitro* model), the uptake of both nanoparticle samples occurred after 5 min of incubation and amounted to 7% and 10% for Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur samples,



Amph-PVP-Cur (amphiphilic derivative-poly-N-vinylpyrrolidonecurcumin)

Amph-PVP-AA-Mal-Cur (amphiphilic derivative-poly-Nvinylpyrrolidone-acrylic acid-maleimide group-curcumin)

Fig. 2. In vitro release profiles of curcumin from the Amph-PVP-Cur and Amph-PVP-AK-Mal-Cur nanoparticles. Free curcumin was used as a control.

respectively. With prolonged incubation, the accumulation levels of both samples increased significantly, reaching 36% and 29% after 30 min and 75% and 78% after 60 min of incubation, respectively.

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For spheroids (3D *in vitro* model), no significant difference in sample accumulation was observed, but the accumulation efficiency decreased significantly. For example, the accumulation levels of Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur were 29% and 34% after 1 h of incubation, 33% and 32% after 3 h, and 59% and 70% after 7 h, respectively.

Study of cytotoxic activity of curcumin-loaded polymeric nanoparticles

The cytotoxicity of Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur curcumin-loaded polymeric nanoparticles was evaluated on the OVCAR-3 ovarian adenocarcinoma cell line using 2D and 3D *in vitro* models with the MTT assay (Table 2, Fig. 4). Empty polymeric nanoparticles (without curcumin), in particular, Amph-PVP, Amph-PVP-AA, and Amph-PVP-AA-Mal samples were used as controls.

For monolayer cell culture, control samples (without curcumin) remained non-toxic even at its maximum concentration after a 48 h-incubation period (IC50 > 500 μ g/mL). Meanwhile, curcumin-loaded samples started to exert cytotoxic effects after 48 h of incubation. Thus, IC50 was 211 ± 13 μ g/mL and 137 ± 9 μ g/mL for Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur samples, respectively. A comparable tendency was noted in tumor spheroids: the Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur samples demonstrated heightened cytoxicity following 48 h (as indicated by the characteristics of the corresponding curves). However, cytoxicity of these samples did not reach the semi-inhibitory concentration (IC50 > 500 μ g/mL).

DISCUSSION

Summary of the primary study outcomes

New data on the effect of nanoparticles based on amphiphilic PVP derivatives and its copolymers with acrylic acid on OVCAR-3 tumor cells were obtained. The study demonstrated that neither the original polymeric

Table 1. Characteristics of pol	ymer nanoparticles
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Particle type	Average size, nm	Polydispersity index	ζ-potential, mV
Amph-PVP-Cur	204	0.269	-18±-2
Amph-PVP-AA-Mal-Cur	352	0.358	-15±-1.5

Note. Amph-PVP-Cur — nanoparticles from amphiphilic derivatives of poly-N-vinylpyrrolidone loaded with curcumin; Amph-PVP-AA-Mal-Cur — nanoparticles from amphiphilic derivatives of poly-N-vinylpyrrolidone copolymers with acrylic acid modified with maleimide and loaded with curcumin.

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Amph-PVP-AA-Mal-Cur (amphiphilic derivative-poly-N-vinylpyrrolidone-acrylic acid-maleimide group-curcumin)

Fig. 3. Accumulation efficiency of the polymeric nanoparticles loaded with curcumin in monolayer culture (2D *in vitro* model) and tumor spheroids (3D *in vitro* model) from human ovarian adenocarcinoma OVCAR-3 cells Flow cytometry (2D *in vitro* model) and fluorimetry (3D *in vitro* model). **** p < 0.001.



Fig. 4. Cytotoxicity of the polymeric nanoparticles in monolayer culture (2D *in vitro* model) and in tumor spheroids (3D *in vitro* model) from human ovarian adenocarcinoma OVCAR-3 cells after incubation for 24 and 48 h. MTT-test.

		IC50*, µg/mL				
Sample	2D in v	2D <i>in vitro</i> model		3D <i>in vitro</i> model		
	24 h	48 h	24 h	48 h		
Amph-PVP-Cur		211±13	> 500			
Amph-PVP-AA-Mal-Cur	> 500	137±9				

Table 2. Cytotoxicity of polymeric nanoparticles in monolayer culture (2D in vitro model) and tumor spheroids (3D in vitro model) from human ovarian adenocarcinoma OVCAR-3 cells. MTT test

Note. * IC50 is a semi-inhibitory concentration. Amph-PVP — nanoparticles from amphiphilic derivatives of poly-N-vinylpyrrolidone; Amph-PVP-AA — nanoparticles from copolymers of amphiphilic derivatives of poly-N-vinylpyrrolidone with acrylic acid; Amph-PVP-AA-Mal — specified nanoparticles modified with maleimide; Amph-PVP-Cur — nanoparticles from amphiphilic derivatives of poly-N-vinylpyrrolidone loaded with curcumin; Amph-PVP-AA-Mal-Cur — nanoparticles from amphiphilic derivatives of poly-N-vinylpyrrolidone copolymers with acrylic acid modified with maleimide and loaded with curcumin.

nanoparticles (Amph-PVP), nor those based on a PVP copolymer with acrylic acid (Amph-PVP-AA), nor the same nanoparticles modified with maleimide (Amph-PVP-AA-Mal) exhibited any signs of toxicity. However, upon loading with curcumin, Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur nanoparticles demonstrated a significant cytotoxic effect in the 2D model and exhibited a weaker yet evident cytotoxic effect on spheroids derived from human ovarian adenocarcinoma OVCAR-3 cells in the 3D model.

Discussion of the primary study outcomes

Novel nanoparticles were developed based on amphiphilic polymers and copolymers of N-vinylpyrrolidone and acrylic acid (Amph-PVP, Amph-PVP-AA) loaded with the lipophilic antitumor agent curcumin. The resulting carrier nanoparticles were characterized by their physicochemical properties (average size and ζ -potential) as well as accumulation efficiency in OVCAR-3 tumor cells. In addition, their cytotoxicity was evaluated in 2D (monolayer culture) and 3D (tumor spheroids) *in vitro* models of OVCAR-3 human ovarian adenocarcinoma.

Multicellular tumor spheroids have recently been considered as a more relevant *in vitro* model compared to monolayer cell culture. Due to their three-dimensional structure, tumor spheroids can mimic the heterogeneity and microenvironment of small solid tumors *in vivo*. This includes specific gene expression, intercellular interactions, as well as cell-cell contacts with the extracellular matrix, growth kinetics, metabolic rate, and resistance to chemotherapy [17].

Furthermore, this study demonstrated that the internalization (uptake) of nanoparticles by cells, whether with the surface modified or unmodified by additional maleimide groups containing curcumin within its hydrophobic core (Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur), exhibited comparable efficiency in both cases. Consequently, the use of nanoparticles derived from PVP copolymer derivatives with acrylic acid modified with maleimide groups did not have any significant effect on the penetration of nanocarriers into cells. In tumor spheroids,

accumulation was much slower than in monolayer culture, which can be explained by a more complex and prolonged penetration of nanocarriers inside the volumetric multilayer spheroid structure. These findings highlight the need to study nanocarriers not only in 2D monolayer cultures, which are currently used, but also in 3D *in vitro* models, which allow a more accurate simulation of solid tumor conditions *in vivo* compared to classical 2D models. In addition, testing with 3D *in vitro* models has an important humanitarian component by reducing the number of animals used in subsequent nonclinical studies.

When the cytotoxicity of the samples was evaluated by the MTT assay, the Amph-PVP-based curcumin-loaded nanoparticles were found to suppress the metabolic activity of human ovarian adenocarcinoma monolayer culture cells after 48 h of incubation. For spheroids, Amph-PVP-Cur and Amph-PVP-AA-Mal-Cur samples also showed a moderate cytotoxic effect after 48 h of incubation without reaching the semi-inhibitory concentration. This result can be explained by the longer time required for both the accumulation of nanoparticles in the cells of multicellular spheroids and the subsequent release of curcumin from the nanoparticles in the spheroids. As anticipated, the cells in the spheroids exhibited increased resistance to the antitumor drug's action compared to the monolayer culture. This finding is well correlated with prior studies involving different cell lines and antitumor agents [9], as well as other nanocarriers, such as polymeric thymoguinone-loaded nanocontainers [18] and doxorubicin-loaded polyelectrolyte capsules [19].

Additionally, the empty polymeric nanoparticles (without curcumin) used as control showed no cytotoxicity in both 2D and 3D *in vitro* models even after 48 h incubation at all concentrations tested, indicating their lack of toxicity.

Study limitations

The study limitations include the insufficient time range where the effect of nanoparticles on tumor cells was examined. However, the cytotoxic effect of curcuminloaded nanoparticles would be more pronounced with an extended incubation period of 72 h or more.

CONCLUSION

In the study, several nanocarriers were obtained based on amphiphilic PVP derivatives and its copolymer with acrylic acid. These nanocarriers included nanoparticles with a surface modified with maleimide functional groups. Such modification of nanoparticles is necessary for their further conjugation with targeted tumor-specific molecules in the development of combined targeted nanosystems with antitumor properties. None of the modifications of the resulting nanoparticles showed toxicity toward human ovarian adenocarcinoma cells. Empty nanocarriers without curcumin loading are non-toxic, thus serving as a foundation for the development of novel and effective delivery systems for antitumor agents, including targeted therapy following the incorporation of specific ligands. The curcumin-loaded nanoparticles was demonstrated to exhibit a pronounced cytotoxic effect in a 2D in vitro model using the OVCAR-3 human ovarian adenocarcinoma cell line. A moderate cytotoxic effect was observed in a 3D in vitro model. These findings suggest that the developed nanoparticles possess considerable potential and could provide a foundation for the development of novel and effective targeting nanosystems for the delivery of antitumor agents with high bioavailability in the future.

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ADDITIONAL INFORMATION

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